



Alt-R™ CRISPR-Cas9 System

| *In vitro* cleavage of target DNA with
| ribonucleoprotein complex

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Revision history

| Version | Release date | Description of changes |
|---------|---------------|---|
| 3 | October 2021 | Corrected concentrations of reagents, updated wording of text. |
| 2.2 | August 2019 | Corrected a component amount needed to create the RNP complex. |
| 2.1 | April 2019 | Adjusted component amount needed to perform the <i>in vitro</i> digestion reaction from 100 nM to 50 nM DNA substrate. |
| 2 | July 2018 | Added instructions for using Alt-R CRISPR-Cas9 sgRNA. Updated names and catalog numbers for Alt-R enzymes (V3). |
| 1.1 | May 2018 | Added note about use of improved Alt-R enzymes (V3): direct substitution in protocol of V3 enzymes for original enzymes (3NLS). |
| 1 | November 2017 | Original publication |

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Introduction

This protocol describes how to use a Cas9 ribonucleoprotein (RNP) complex to enable *in vitro* cleavage of double-stranded, targeted DNA. The Cas9 RNP complex contains both an Alt-R CRISPR-Cas9 guide RNA (crRNA:tracrRNA duplex or sgRNA) and an *S. pyogenes* Cas9 endonuclease. This protocol demonstrates a method to experimentally confirm the activity of CRISPR guide RNA before practical application.

Consumables

| Reagents | Ordering information |
|--|---|
| Option 1, 2-part guide RNA (crRNA + tracrRNA): | |
| <ul style="list-style-type: none">Alt-R CRISPR-Cas9 crRNA or Alt-R CRISPR-Cas9 crRNA XT | IDT predesigned and custom crRNA (www.idtdna.com/CRISPR-Cas9) |
| <ul style="list-style-type: none">Alt-R CRISPR-Cas9 tracrRNA or Alt-R CRISPR-Cas9 tracrRNA – ATTO™ 550 | IDT (cat # 1072532, 1072533, 1072534) IDT (cat # 1075927, 1075928) |
| Option 2, single guide RNA (sgRNA): Alt-R CRISPR-Cas9 sgRNA | IDT predesigned and custom sgRNA: (www.idtdna.com/CRISPR-Cas9) |
| Alt-R <i>S.p.</i> Cas9 Nuclease V3 | IDT (cat # 1081058, 1081059) |
| Alternative: Alt-R <i>S.p.</i> HiFi Cas9 Nuclease V3 | IDT (cat # 1081060, 1081061) |
| Nuclease-Free Duplex Buffer | IDT (cat # 11-05-01-12, various sizes available) |
| Nuclease-Free IDTE, pH 7.5 (1X TE solution) | IDT (cat # 11-01-02-02) |
| Nuclease-Free Water | IDT (cat # 11-04-02-01) |
| DNA substrate containing the target sequence | gBlocks Gene Fragments (www.idtdna.com/gBlocks), or similar |
| For 10X Cas9 Nuclease Reaction Buffer combine: | |
| <ul style="list-style-type: none">200 mM HEPES1 M NaCl50 mM MgCl₂1 mM EDTA, pH 6.5 at 25°C | General laboratory supplier |
| PBS | |
| Alternative: For Cas9 Dilution Buffer, combine: | General laboratory supplier |
| <ul style="list-style-type: none">30 mM HEPES150 mM KCl, pH 7.5 | |
| Proteinase K (Molecular biology grade) | General laboratory supplier |

Protocol

Prepare the double-stranded DNA template as cleavage substrate

Design your template, considering the following:

- Multiple types of double-stranded DNA can be used as substrates for Cas9 cleavage. Three common examples are:
 - linearized plasmid
 - purified PCR products
 - duplexed synthetic oligos
- Your template must contain a 20 nt guide sequence, followed by the Cas9 PAM site (NGG).
- The guide sequence should match the target-specific guide RNA that will be used in **Perform the *in vitro* digestion reaction**.
- The amount of template needed for the digestion may vary depending on the detection method used to **Visualize cleaved products** and the template size, as shown in this table:

| DNA template | Length (bp) | Final concentration | Visualization method |
|---------------------------------------|-------------|---------------------|--|
| DNA oligo duplex | 30–100 | 2–5 μ M | PAGE |
| gBlocks™ Gene Fragment or PCR product | 100–500 | 5–50 nM | Fragment Analyzer (Agilent), agarose gel |
| gBlocks™ Gene Fragment or PCR product | 500–2000 | 2–5 nM | |
| Linearized plasmid | \geq 2000 | 1–2 nM | |

1. Make sure you are using a 10:1 molar ratio of Cas9 RNP:DNA substrate to obtain the best cleavage efficiency.
2. Resuspend or dilute the DNA substrate in Nuclease-Free Water to the required concentration.



Note: IDT provides a resuspension calculator at www.idtdna.com/SciTools.

Prepare the guide RNA

1. Resuspend each RNA oligo (Alt-R CRISPR-Cas9 crRNA, tracrRNA, sgRNA) in IDTE buffer to a stock concentration of 100 μM .
2. If you are using sgRNA, dilute it to **10 μM** , then dilute it to a working concentration of 10 μM (1:10 dilution) in IDTE Buffer before proceeding to the next section: **Create the RNP complex**.
3. Mix the crRNA and tracrRNA oligos in equimolar concentrations in a sterile microcentrifuge tube to a final duplex concentration of 10 μM . The following table shows an example of a 10 μL final volume duplex:

| Component | Amount (μL) |
|--|--------------------------|
| 100 μM Alt-R CRISPR-Cas9 crRNA | 1 |
| 100 μM Alt-R CRISPR-Cas9 tracrRNA | 1 |
| Nuclease-Free Duplex Buffer | 8 |
| Total volume | 10 |

4. Heat the duplex at 95°C for 5 min.
5. Remove from heat and allow to cool to room temperature (15–25°C).

Create the RNP complex

1. Combine the guide RNA and Cas9 enzyme in equimolar amounts.

| Component | Amount (μL) |
|---|--------------------------|
| 10 μM Alt-R guide RNA [From Prepare the guide RNA , step 2 (sgRNA), or step 5 (crRNA:tracrRNA)] | 10 |
| Alt-R <i>S.p.</i> Cas9 enzyme (62 μM stock)* | 1.6 |
| PBS† | 88.4 |
| Total volume | 100 |

* All Alt-R *S.p.* Cas9 enzymes are provided at a stock concentration of 62 μM .

† Cas9 RNP complexes can be made in PBS or in Cas9 dilution buffer (30 mM HEPES, 150 mM KCl, pH 7.5).

2. Incubate at room temperature for 5–10 min for optimal formation of the RNP complex.

Perform the *in vitro* digestion reaction

1. Assemble the reaction at room temperature (15–25°C).

| Component | Amount (μL) |
|-----------------------------------|-------------|
| 10X Cas9 Nuclease Reaction Buffer | 1 |
| 1 μM Cas9 RNP | 1 |
| 100 nM DNA substrate | 1 |
| Nuclease-Free Water | 7 |
| Total volume | 10 |

2. Incubate the reaction at 37°C for 60 min.
3. Add 1 μL Proteinase K (20 mg/mL) to the reaction, then incubate the mixture at 56°C for 10 min to release the DNA substrate from the Cas9 endonuclease.

Visualize cleaved products

Analyze the digestion by using one of the following methods:

- Agarose gel electrophoresis
- Fragment Analyzer™ System (Agilent), or similar

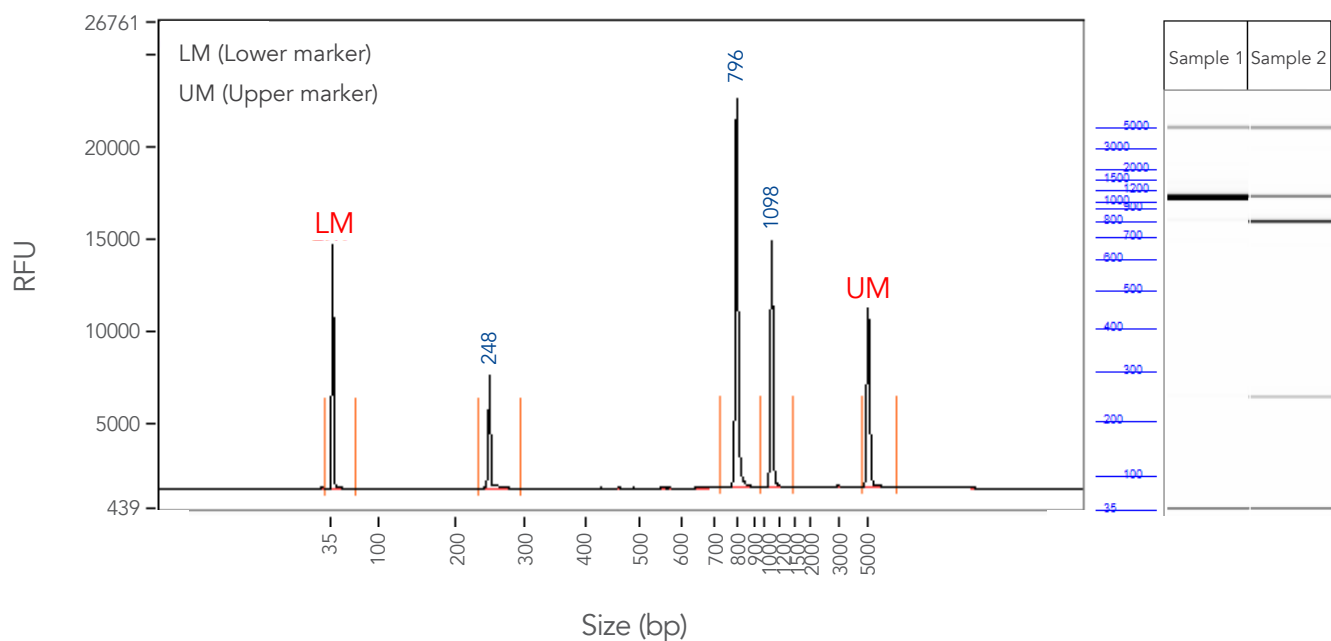


Figure 1. Sample data showing *in vitro* digestion reaction using Alt-R tracrRNA with Alt-R CRISPR-Cas9 Positive Control crRNA (HPRT). Column-purified PCR product consisting of the Hs HPRT crRNA positive control sequence was used as a template in a 10 μL *in vitro* Cas9 digestion reaction. Sample 1 contains template without RNP. Sample 2 contains template and RNP. Digestion reactions were analyzed on a Fragment Analyzer™ system and a gel imaging system. Trace (left) shows results from Sample 2. Gel image (right) shows results for Samples 1 and 2.

Alt-R CRISPR-Cas9 System: *In vitro* cleavage of target DNA with RNP complex

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