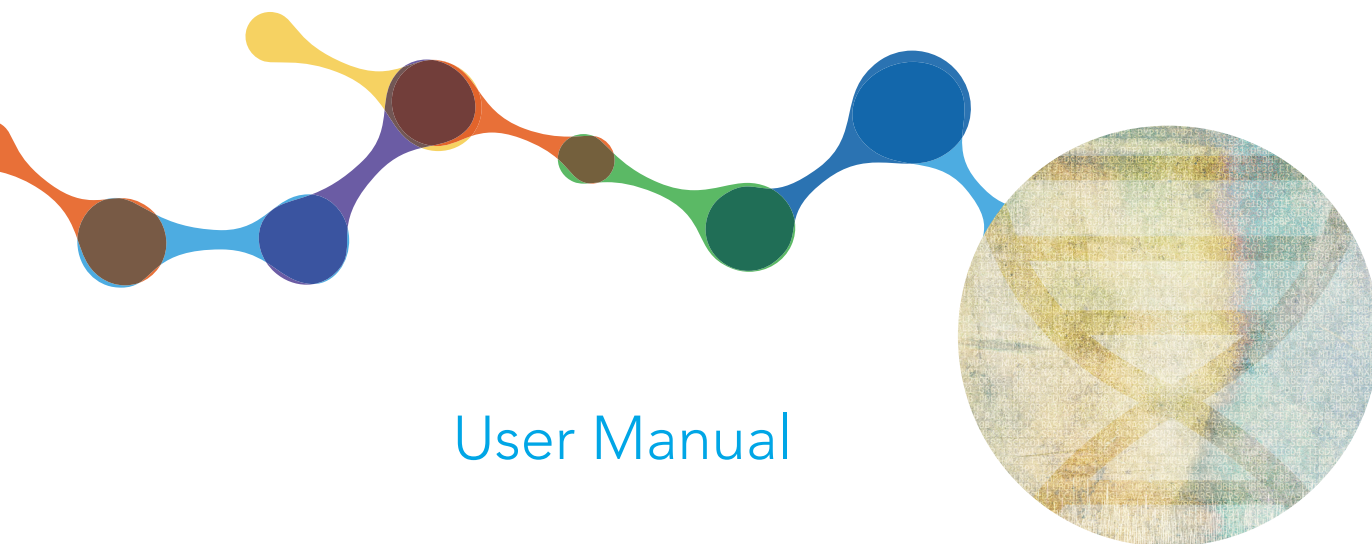


# DNaseAlert™ Substrate Nuclease Detection System



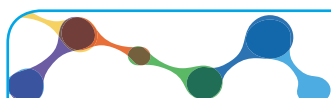
User Manual

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## Introduction

Nucleases are widely present in the laboratory environment and can interfere with many experiments. IDT developed DNaseAlert™ and RNaseAlert® Substrate Nuclease Detection Systems for rapid, sensitive detection of DNases and RNases, respectively. These reagents are fluorescence-quenched oligonucleotide probes that fluoresce after nuclease degradation. The assays can be read visually or measured and quantified using a fluorometer. Assays can be used qualitatively or quantitatively to test lab reagents, work surfaces, equipment, and supplies for nuclease contamination. The reagents can also be used as highly sensitive substrates for enzyme kinetic studies.

The DNaseAlert Substrate is a synthetic DNA oligonucleotide that has a HEX™ reporter dye (hexachlorofluorescein, R) on one end and a dark quencher (Q) on the other end (Figure 1). Its sequence has been carefully optimized to react with a wide variety of nucleases; it contains domains that will react with single-stranded endonucleases, single-stranded exonucleases, and double-stranded nucleases. Intact, the substrate has little or no fluorescence. When cleaved by a DNase, the substrate fluoresces pink (536 nm or UV excitation, 556 nm emission).

The DNaseAlert test is fast and simple. Lab surfaces or liquid reagents can be tested and verified as being “nuclease-free” or “contaminated” in less than one hour. For speed and ease of use, a simple tube-based visual assay can be performed directly at the site of testing. Alternatively, quantitative fluorescent results (cuvette or microtiter plate formats) can be obtained and used to document nuclease testing for GLP or manufacturing needs.

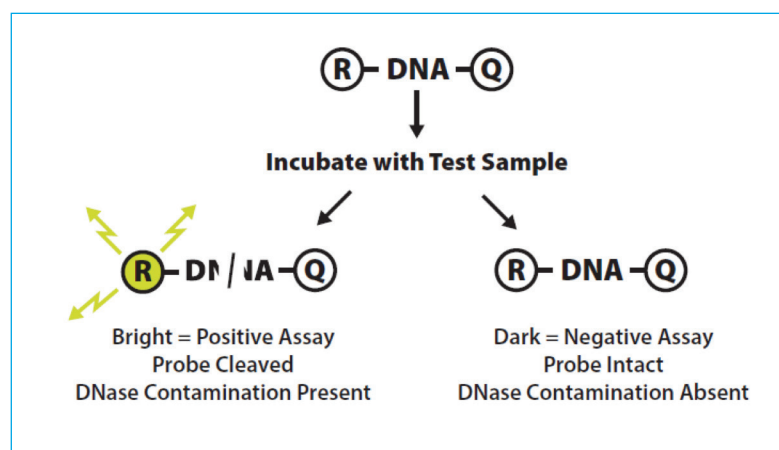


Figure 1. DNaseAlert™ Assay.

DNaseAlert™  
Substrate Nuclease  
Detection System

## Kit Components and Storage Conditions

Component	Quantity	Storage Condition*
DNaseAlert™ Substrate	25 single-use tubes (50 pmol per tube) <b>or</b> 2 tubes bulk substrate (2 nmol per tube)	-20°C (protect from light to prevent photobleaching)
DNaseAlert Buffer	250 µL	-20°C
Nuclease-Free Water	2 mL	Room temperature
DNase I Enzyme (positive control)	25 µL	-20°C
Nuclease Decontamination Solution	50 mL	Room temperature

\* Reagent quality is guaranteed for 6 months from date received when stored as directed.



### Before you start:

- Use gloves when handling kit components and performing the assay.
- Perform all steps under nuclease-free conditions:
  - Use nuclease-free pipette tips and tubes.
  - If necessary, clean pipettes and other lab surfaces with Nuclease Decontamination Solution before use.



## Single-use tubes—testing a liquid solution

Set up one DNaseAlert assay for each sample to be tested. Include 2 additional tubes for the positive and negative controls.

1. Add 5 µL of Nuclease-Free Water (provided) to a DNaseAlert Substrate single-use tube for each assay.
2. Add 5 µL of 10X DNaseAlert Buffer to each tube.
3. Add 40 µL of test sample to each sample tube. Add 40 µL of Nuclease-Free Water to each control tube. Add 1 µL of DNase I to the positive control tube. **Mix thoroughly.** The final concentration of substrate is 1 µM for the visual assay; lower substrate concentrations can be used for fluorometric detection (see **Quantitative measurement**, below).
4. **Incubate at 37°C for 30–60 min.** Greater sensitivity is achieved with longer incubations. If a temperature-regulated incubator or water bath is not available, incubation can be done at room temperature (2–3X longer incubation time is recommended to achieve similar sensitivity).

### 5. Perform detection.

**Quick visual confirmation:** Place tube on a shortwave (300 nm) UV transilluminator. For best results, use a darkened room.

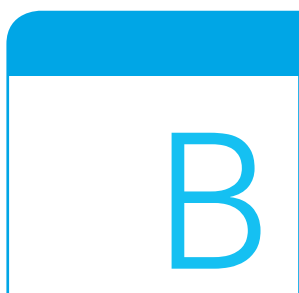
If the tube remains clear, the assay is negative and the sample is free of detectable DNase contamination. If the tube glows pink, DNase contamination is present. A longwave (365 nm) UV source can be used, but sensitivity will be lower.



**Important safety note:** Never look directly into a UV light source. Always use protective face shielding.

**Quantitative measurement:** Place contents in a nuclease-free cuvette or microtiter plate. Measure fluorescence using a fluorometer set to the HEX channel (536 nm excitation, 556 nm emission). After the incubation at Step 4 (above) is complete, the sample can be diluted with up to 2 mL Nuclease-Free Water, as needed, to accommodate the size of the detection chamber.

6. Validate all negative assays. Add 1  $\mu\text{L}$  of DNase I (or other DNase) to each tube. Mix and incubate at 37°C for 10 min. Repeat Step 5 (Perform detection). All negative tubes should now be positive. Any assays that fail to fluoresce during this validation step must be considered “failed”, and the nuclease detection assay should be repeated. See troubleshooting guide at the end of this manual for help with assay failures.

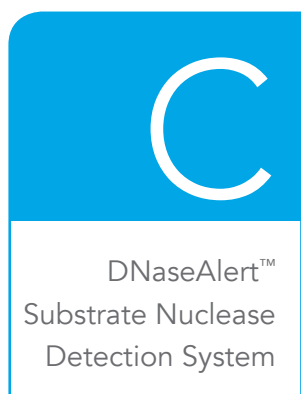


## Single-use tubes—testing a solid object or dry surface

Set up one DNaseAlert assay for each sample to be tested. Include 2 additional tubes for the positive and negative controls.

1. Add 5  $\mu\text{L}$  of Nuclease-Free Water (provided) to a DNaseAlert Substrate single-use tube for each assay.
2. Add 5  $\mu\text{L}$  of 10X DNaseAlert Buffer to each tube.
3. Add 40  $\mu\text{L}$  of Nuclease-Free Water to each tube. Mix thoroughly.
4. Place the object to be tested into a sample tube. Add 1  $\mu\text{L}$  of DNase I to the positive control tube.
  - Pipette tips, electrodes, or other small solids can be dipped directly into the assay solution.
  - To test surfaces that cannot be dipped into an assay tube:
    - a. Wipe the surface of interest with a piece of nuclease-free filter paper pre-wetted with Nuclease-Free Water.
    - b. Soak the filter paper in a small amount of Nuclease-Free Water.
    - c. Transfer the liquid to an assay tube and proceed as instructed in A. Single-use tubes—testing liquid samples.

5. Incubate at 37°C for 30–60 min.
6. Read assay in a fluorometer using the HEX channel (536 nm excitation, 556 nm emission). After Step 5 incubation is complete, the sample can be diluted using up to 2 mL Nuclease-Free Water, as needed, to accommodate the size of the detection chamber.
7. Validate all negative assays. Add 1  $\mu\text{L}$  of DNase I (or other DNase) to each negative (non-fluorescent) tube. Mix and incubate at 37°C for 10 min. Repeat detection procedure as before. All negative tubes should now be positive. Any assays that fail to fluoresce during this validation step must be considered “failed”, and the nuclease detection assay should be repeated. See troubleshooting guide at the end of this manual for help with assay failures.



## Bulk substrate—testing a liquid solution

Bulk substrate assays can be performed in individual tubes; however, it is more convenient to perform the assays in microtiter plates. We recommend using black opaque plates that minimize scatter and cross-talk between wells.

Always include negative and positive control wells. Use duplicate or triplicate sample wells for quantitative measurements.

1. Add 1 mL of Nuclease-Free Water (provided) to the bulk DNaseAlert Substrate tube. Mix thoroughly. Final substrate concentration is 2  $\mu\text{M}$  (20 pmol substrate in 10  $\mu\text{L}$ ).
2. Add 10  $\mu\text{L}$  of DNaseAlert Substrate to each sample and control well.
3. Add 10  $\mu\text{L}$  of 10X DNaseAlert Buffer to each well.
4. Add 80  $\mu\text{L}$  of sample to the sample wells. Add 80  $\mu\text{L}$  of Nuclease-Free Water to each control well. Add 1  $\mu\text{L}$  of DNase I to the positive control well. Mix thoroughly.
5. Incubate at 37°C for 30–60 min. The recommended final concentration of substrate is 200 nM for a standard 96-well fluorometer; more dilute solutions can be used for sensitive cuvette fluorometers.
6. Read assay in fluorometer using the HEX channel (536 nm excitation, 556 nm emission). Assays can be read as simple endpoint measurements or examined in real time to obtain quantitative kinetic curves.
7. Validate all negative assays. Add 1  $\mu\text{L}$  of DNase I (or other DNase) to each well. Mix and incubate at 37°C for 10 min. Repeat detection procedure as before. All negative tubes should now be positive. Any assays that fail to fluoresce during this validation step must be considered “failed”, and the nuclease detection assay should be repeated. See troubleshooting guide at the end of this manual for help with assay failures.

## Troubleshooting

Observation	Possible Problem	Solution
False Negatives	<p><b>Presence of DNase inhibitors</b> Solutions with extreme pH, strong ionic strength, or detergents can block action of DNases, preventing detection of contaminants that are present.</p>	<p><b>Test your solution for inhibitors of the DNaseAlert assay.</b> Set up a standard assay with your test solution and add 1 µL DNase I (or other DNase).  If the assay does not convert to “positive” after a 1-hour incubation, your test solution is incompatible with the DNaseAlert system.</p>
	<p><b>Low pH solutions</b> Solutions with pH &lt;7.0 will decrease the efficiency of HEX fluorescence, lowering assay sensitivity.</p>	<p>If possible, adjust the pH of the test solution to obtain pH &gt;7.0 and &lt;9.0.</p>
	<p><b>Substrate loss</b> 1. The DNaseAlert Substrate is provided dried down. The dry pellet can become dislodged from the bottom of the tube and may be lost from the tube when opened; dry oligos can be electrostatically attracted to laboratory gloves. 2. Prolonged exposure of the DNaseAlert Substrate to light can lead to photobleaching of the fluorescent dye and decrease assay sensitivity.</p>	<p>1. a. <b>Spin down tubes before opening.</b> b. <b>Perform a positive control assay.</b> Always perform a positive control test on single-use tube assays with negative results. A positive result in another tube, even from the same kit, is not sufficient. 2. <b>Store bulk substrate and assay tubes in the dark.</b></p>
	<p><b>Nuclease–substrate incompatibility</b> The DNaseAlert Substrate contains a fluorescent dye at the 5' end and a fluorescence quencher at the 3' end. Activity of some exonucleases is blocked by terminal end-groups and, therefore, such nucleases (e.g., <i>E. coli</i> Exo I) cannot be detected using this assay.</p>	
False Positives	<p><b>Contamination</b> Nuclease contamination of tubes, pipette tips, and other lab equipment can lead to false positives.</p>	<p><b>Perform a negative control assay.</b> A negative control must be included with each set of assays performed. Fluorescence in the negative control tube (Nuclease-Free Water test) indicates contamination in the experimental setup  <b>Obtain new supplies and clean work area.</b> If contamination is suspected, obtain fresh tubes and pipette tips and clean all lab surfaces with Nuclease Decontamination Solution before repeating the assay.</p>
	<p><b>Quencher exhaustion</b> Prolonged exposure of the substrate to UV light can damage the quencher. A visual assay left on an intense shortwave (254 nm) UV source can turn positive even when no nuclease is present.</p>	<p><b>Read your assay immediately to limit exposure to UV light.</b></p>
	<p><b>Substrate degradation</b> The DNaseAlert Substrate contains DNA bases that will be depurinated and degraded when exposed to acidic conditions.</p>	<p><b>Avoid use of solutions with pH &lt;4.5.</b></p>
	<p><b>Substrate stimulation</b> Certain organic solvents disrupt quenching. In particular, DNaseAlert Substrate always glows in acetonitrile.</p>	